

ISSN 2518-1629 (Online),
ISSN 2224-5308 (Print)

ҚАЗАҚСТАН РЕСПУБЛИКАСЫ
ҰЛТТЫҚ ҒЫЛЫМ АКАДЕМИЯСЫНЫҢ
С. Ж. Асфендияров атындағы Қазақ ұлттық медицина университеті

Х А Б А Р Л А Р Ы

ИЗВЕСТИЯ

НАЦИОНАЛЬНОЙ АКАДЕМИИ НАУК
РЕСПУБЛИКИ КАЗАХСТАН
Казакский национальный медицинский
университет им. С. Д. Асфендиярова

NEWS

OF THE NATIONAL ACADEMY OF SCIENCES
OF THE REPUBLIC OF KAZAKHSTAN
Asfendiyarov
Kazakh National Medical University

**SERIES
OF BIOLOGICAL AND MEDICAL**

2 (338)

MARCH – APRIL 2020

PUBLISHED SINCE JANUARY 1963

PUBLISHED 6 TIMES A YEAR

ALMATY, NAS RK

Б а с р е д а к т о р

ҚР ҰҒА академигі, м. ғ. д., проф.

Ж. А. Арзықұлов

Абжанов Архат, проф. (Бостон, АҚШ),
Абелев С.К., проф. (Мәскеу, Ресей),
Айтқожина Н.А., проф., академик (Қазақстан)
Акшулаков С.К., проф., академик (Қазақстан)
Алшынбаев М.К., проф., академик (Қазақстан)
Бәтпенев Н.Д., проф., корр.-мүшесі (Қазақстан)
Березин В.Э., проф., корр.-мүшесі (Қазақстан)
Берсімбаев Р.И., проф., академик (Қазақстан)
Беркінбаев С.Ф., проф., (Қазақстан)
Бисенбаев А.К., проф., академик (Қазақстан)
Бишимбаева Н.Қ., проф., академик (Қазақстан)
Ботабекова Т.К., проф., корр.-мүшесі (Қазақстан)
Bosch Ernesto, prof. (Spain)
Давлетов Қ.К., ассоц.проф., жауапты хатшы
Жансүгірова Л.Б., б.ғ.к., проф. (Қазақстан)
Ellenbogen Adrian, prof. (Tel-Aviv, Israel),
Жамбакин Қ.Ж., проф., академик (Қазақстан), бас ред. орынбасары
Заядан Б.К., проф., корр.-мүшесі (Қазақстан)
Ishchenko Alexander, prof. (Villejuif, France)
Исаева Р.Б., проф., (Қазақстан)
Кайдарова Д.Р., проф., академик (Қазақстан)
Кохметова А.М., проф., корр.-мүшесі (Қазақстан)
Күзденбаева Р.С., проф., академик (Қазақстан)
Локшин В.Н., проф., корр.-мүшесі (Қазақстан)
Лось Д.А., prof. (Мәскеу, Ресей)
Lunenfeld Bruno, prof. (Израиль)
Макашев Е.К., проф., корр.-мүшесі (Қазақстан)
Миталипов Ш.М., (Америка)
Муминов Т.А., проф., академик (Қазақстан)
Огарь Н.П., проф., корр.-мүшесі (Қазақстан)
Омаров Р.Т., б.ғ.к., проф., (Қазақстан)
Продеус А.П., проф. (Ресей)
Purton Saul, prof. (London, UK)
Рахыпбеков Т.К., проф., корр.-мүшесі (Қазақстан)
Сапарбаев Мұрат, проф. (Париж, Франция)
Сарбасов Дос, проф. (Хьюстон, АҚШ)
Тұрысбеков Е.К., б.ғ.к., асс.проф. (Қазақстан)
Шарманов А.Т., проф. (АҚШ)

«ҚР ҰҒА Хабарлары. Биология және медициналық сериясы».

ISSN 2518-1629 (Online), ISSN 2224-5308 (Print)

Меншіктенуші: «Қазақстан Республикасының Ұлттық ғылым академиясы» РҚБ (Алматы қ.).

Қазақстан республикасының Мәдениет пен ақпарат министрлігінің Ақпарат және мұрағат комитетінде
01.06.2006 ж. берілген №5546-Ж мерзімдік басылым тіркеуіне қойылу туралы куәлік.

Мерзімділігі: жылына 6 рет.

Тиражы: 300 дана.

Редакцияның мекенжайы: 050010, Алматы қ., Шевченко көш., 28; 219, 220 бөл.; тел.: 272-13-19, 272-13-18;
<http://biological-medical.kz/index.php/en/>

© Қазақстан Республикасының Ұлттық ғылым академиясы, 2020

Типографияның мекенжайы: «NurNaz GRACE», Алматы қ., Рысқұлов көш., 103.

Г л а в н ы й р е д а к т о р
академик НАН РК, д.м.н., проф.
Ж. А. Арзыкулов

Абжанов Архат, проф. (Бостон, США),
Абелев С.К., проф. (Москва, Россия),
Айтхожина Н.А., проф., академик (Казахстан)
Акшулаков С.К., проф., академик (Казахстан)
Алчинбаев М.К., проф., академик (Казахстан)
Батпенов Н.Д., проф., чл.-корр. (Казахстан)
Березин В.Э., проф., чл.-корр. (Казахстан)
Берсимбаев Р.И., проф., академик (Казахстан)
Беркинбаев С.Ф., проф. (Казахстан)
Бисенбаев А.К., проф., академик (Казахстан)
Бишимбаева Н.К., проф., академик (Казахстан)
Ботабекова Т.К., проф., чл.-корр. (Казахстан)
Bosch Ernesto, prof. (Spain)
Давлетов К.К., ассоц. проф., ответственный секретарь
Джансугурова Л. Б., к.б.н., проф. (Казахстан)
Ellenbogen Adrian, prof. (Tel-Aviv, Israel),
Жамбакин К.Ж., проф., академик (Казахстан), зам. гл. ред.
Заядан Б.К., проф., чл.-корр. (Казахстан)
Ishchenko Alexander, prof. (Villejuif, France)
Исаева Р.Б., проф. (Казахстан)
Кайдарова Д.Р., проф., академик (Казахстан)
Кохметова А.М., проф., чл.-корр. (Казахстан)
Кузденбаева Р.С., проф., академик (Казахстан)
Локшин В.Н., проф., чл.-корр. (Казахстан)
Лось Д.А., prof. (Москва, Россия)
Lunenfeld Bruno, prof. (Израиль)
Макашев Е.К., проф., чл.-корр. (Казахстан)
Миталипов Ш.М., (Америка)
Муминов Т.А., проф., академик (Казахстан)
Огарь Н.П., проф., чл.-корр. (Казахстан)
Омаров Р.Т., к.б.н., проф. (Казахстан)
Продеус А.П., проф. (Россия)
Purton Saul, prof. (London, UK)
Рахыпбеков Т.К., проф., чл.-корр. (Казахстан)
Сапарбаев Мурат, проф. (Париж, Франция)
Сарбасов Дос, проф. (Хьюстон, США)
Турысбеков Е. К., к.б.н., асс. проф. (Казахстан)
Шарманов А.Т., проф. (США)

«Известия НАН РК. Серия биологическая и медицинская».

ISSN 2518-1629 (Online), ISSN 2224-5308 (Print)

Собственник: РОО «Национальная академия наук Республики Казахстан» (г. Алматы).

Свидетельство о постановке на учет периодического печатного издания в Комитете информации и архивов Министерства культуры и информации Республики Казахстан №5546-Ж, выданное 01.06.2006 г.

Периодичность: 6 раз в год.

Тираж: 300 экземпляров.

Адрес редакции: 050010, г. Алматы, ул. Шевченко, 28; ком. 219, 220; тел. 272-13-19, 272-13-18;

<http://biological-medical.kz/index.php/en/>

© Национальная академия наук Республики Казахстан, 2020

Адрес типографии: «NurNazGRACE», г. Алматы, ул. Рыскулова, 103.

Editor in chief

Zh.A. Arzykulov,
academician of NAS RK, Dr. med., prof.

Abzhanov Arkhat, prof. (Boston, USA),
Abelev S.K., prof. (Moscow, Russia),
Aitkhozhina N.A., prof., academician (Kazakhstan)
Akshulakov S.K., prof., academician (Kazakhstan)
Alchinbayev M.K., prof., academician (Kazakhstan)
Batpenov N.D., prof., corr. member (Kazakhstan)
Berezin V.Ye., prof., corr. member (Kazakhstan)
Bersimbayev R.I., prof., academician (Kazakhstan)
Berkinbaev S.F., prof. (Kazakhstan)
Bisenbayev A.K., prof., academician (Kazakhstan)
Bishimbayeva N.K., prof., academician (Kazakhstan)
Botabekova T.K., prof., corr. member (Kazakhstan)
Bosch Ernesto, prof. (Spain)
Davletov Kairat, PhD, associate professor, executive Secretary
Dzhansugurova L.B., Cand. biol., prof. (Kazakhstan)
Ellenbogen Adrian, prof. (Tel-Aviv, Israel),
Zhambakin K.Zh., prof., academician (Kazakhstan), deputy editor-in-chief
Ishchenko Alexander, prof. (Villejuif, France)
Isayeva R.B., prof. (Kazakhstan)
Kaydarova D.R., prof., academician (Kazakhstan)
Kokhmetova A., prof., corr. member (Kazakhstan)
Kuzdenbayeva R.S., prof., academician (Kazakhstan)
Lokshin V.N., prof., corr. member (Kazakhstan)
Los D.A., prof. (Moscow, Russia)
Lunefeld Bruno, prof. (Israel)
Makashev E.K., prof., corr. member (Kazakhstan)
Mitalipov Sh.M. (America)
Muminov T.A., prof., academician (Kazakhstan)
Ogar N.P., prof., corr. member (Kazakhstan)
Omarov R.T., cand. biol., prof. (Kazakhstan)
Prodeus A.P., prof. (Russia)
Purton Saul, prof. (London, UK)
Rakhypbekov T.K., prof., corr. member (Kazakhstan)
Saparbayev Murat, prof. (Paris, France)
Sarbasov Dos, prof. (Houston, USA)
Turysbekov E.K., cand. biol., assoc. prof. (Kazakhstan)
Sharmanov A.T., prof. (USA)

News of the National Academy of Sciences of the Republic of Kazakhstan. Series of biology and medicine.
ISSN 2518-1629 (Online), ISSN 2224-5308 (Print)

Owner: RPA "National Academy of Sciences of the Republic of Kazakhstan" (Almaty).

The certificate of registration of a periodic printed publication in the Committee of information and archives of the Ministry of culture and information of the Republic of Kazakhstan N 5546-Ж, issued 01.06.2006.

Periodicity: 6 times a year.

Circulation: 300 copies.

Editorial address: 28, Shevchenko str. of. 219, 220, Almaty, 050010; tel. 272-13-19, 272-13-18;

<http://biological-medical.kz/index.php/en/>

© National Academy of Sciences of the Republic of Kazakhstan, 2020

Address of printing house: «NurNaz GRACE», 103, Ryskulov str, Almaty.

NEWS

OF THE NATIONAL ACADEMY OF SCIENCES OF THE REPUBLIC OF KAZAKHSTAN

SERIES OF BIOLOGICAL AND MEDICAL

ISSN 2224-5308

Volume 2, Number 338 (2020), 20 – 30

<https://doi.org/10.32014/2020.2519-1629.9>

UDC 606:577.112; 604.4:577.112.4/8

IRSTI 62.99.31, 62.13.31

**E. T. Tailakova, S. O. Sadikaliyeva, G. O. Shynybekova,
A. K. Abubakirova, K. T. Sultankulova, O. V. Chervyakova**

Research Institute for Biological Safety Problems, Gvardeiskiy, Korday district, Zhambyl region, Kazakhstan.

E-mail: tailakova_86@mail.ru

CONSTRUCTION, EXPRESSION AND PURIFICATION OF *BRUCELLA* *SPP.* RECOMBINANT PROTEINS L7/L12 AND SODC IN *E. COLI*

Abstract. Brucellosis is still an important public health problem as long as natural reservoirs of infection exist. Currently, live attenuated vaccines based on strains S19, RB51 and Rev1 are used for the prevention of brucellosis in animals, the main disadvantage of which is virulence for humans. However, animal immunization programs should be implemented to reduce the incidence of humans. The development of safe and effective new generation vaccines using “omix” technology is a promising direction of vaccinology. A number of immunogenic *Brucella* proteins that elicit both a humoral and cellular immune response has been identified. The aim of these research was to optimize the expression and purification conditions of the *Brucella spp.* recombinant proteins L7/L12 and SodC. As a result, expressing plasmids pET/Br-L7/L12 and pET/Br-SodC were obtained. The parameters of target genes expression in *E. coli* were established and the method for purification of recombinant proteins was optimized. Purification of the L7/L12 protein was performed under hybrid conditions on HisPur agarose using a binding buffer containing 6 M guanidine hydrochloride, a wash buffer with 20 mM imidazole and an elution buffer with 300 mM imidazole. Protein SodC was purified under denaturing conditions with the addition of 1 % Triton X-100 and 1 % sodium deoxycholate to the lysis buffer. Inclusions were solubilized with a buffer containing 8 M urea and 5 mM imidazole. The target protein was eluted from HisPur agarose with buffer containing 8 M urea and 100 mM imidazole. The use of modified purification protocols made it possible to obtain purified recombinant proteins with a yield of 13 mg/L for the L7/L12 protein and 10 mg/L for the protein SodC, respectively. The specificity of the proteins was confirmed by a Western blot. Immunization of mice with recombinant proteins led to the production of specific antibodies, the titer of which in ELISA was 1:20480 and 1:20480, respectively.

Key words: *Brucella spp.*, ribosomal protein L7/L12, superoxide dismutase, expression, protein purification.

Introduction. Brucellosis is one of the widespread zoonotic diseases causing great economic damage to agriculture. According to the Joint FAO Expert Committee, brucellosis of livestock is prevalent in almost the whole world and since animals with brucellosis are a source of infection for humans, this disease is a high degree of danger [1-4]. The annual identification of farm animals and people reacting to brucellosis in certain regions of Kazakhstan indicates an extremely unstable situation for this infection and the real possibility of forming foci of brucellosis with varying degrees of activity of manifestations of epizootic and epidemic processes in farms [5,6]. In this regard, this infection is a serious problem for veterinary and medical science.

Today, recombinant proteins having antigenic and immunogenic properties are widely used in the development of prophylactic and diagnostic preparations, as well as candidates for vaccine against various zoonotic diseases. A lot of work has been devoted to the study of immunogenic proteins of *Brucella spp* and a number of proteins have been discovered, such as: L7/L12, SodC, BP26, BCSP31, Omp16, Omp19, Omp31, which have immunogenic properties that can be used for diagnostic purposes. These proteins are conserved and their identity is 100 % between *Brucella* species [7-12].

In the study of *Brucella* antigens capable of inducing cellular immunity, a 12 kDa protein was detected that causes lymphocyte proliferation and is a L7/L12 ribosomal protein. It was shown that purified recombinant L7/L12 protein produced in *E. coli* stimulates CD4 T cell immunity in mice infected with *B. abortus*. It was also shown that immunization of mice with the recombinant ribosomal protein L7/L12 protects them from control infection with *B. abortus*. According to published data, L7/L12 is an immunodominant *B. abortus* protein that elicits a cellular immune response (Th1 and CD8 + T cells) [13-15].

The periplasmic protein SodC (Cu-Zn superoxide dismutase) is one of the main enzymes of the antioxidant system of microorganisms, which is considered as one of the universal mechanisms of the pathogenesis of infectious diseases, and indicators reflecting shifts in the levels of antioxidant enzymes are key factors in predicting the outcome of the disease. The researchers determined the protective effect of the antioxidant enzyme SodC *B. abortus*, expressed in significant induction of T-cell proliferation and production of gamma-interferon in infected mice. Thus, vaccination of mice with *E. coli* cells expressing the SodC *B. abortus* enzyme formed a defense against brucella infection. The use for this purpose of plasmid DNA, including the SodC gene *B. abortus*, also induced a humoral and cellular immune response against the causative agent of brucellosis [8,16].

The purpose of these studies was the construction of expression vectors, the expression and optimization of the purification conditions of the recombinant *Brucella spp* L7/L12 and SodC proteins, as well as the production of specific sera to them. Recombinant proteins and their specific sera will be used in the development of a vector anti-brucellosis vaccine based on sheep pox virus.

Materials and methods. Bacterial strains. The studies used the vaccine strain *B. abortus* S19 obtained from the laboratory of the collection of microorganisms of the Research Institute of Biological Safety Problems RK ME&S – Science Committee, Kazakhstan. *B. abortus* S19 genomic DNA was isolated using the PrepMan Ultra kit (Applied Biosystems, USA). For manipulation with plasmid DNA, *E. coli* strain TOP10 (Invitrogen, USA) was used. For bacterial expression, *E. coli* T7 express strain (New England Biolabs, USA) was used.

Construction of expression cassettes and obtaining producer strains. The *Brucella* genes L7/L12 and *sodC* were amplified with the genomic DNA *B. abortus* S19 using pairs of primers FP-L7/L12-5'-CGCATATGGCTGATCTCGAAAGATCGT-3', RP-L7/L12-5'-CGCTCGAGCTTGA GTTCAACCTTGGCGCCA-3' and FP-SodC-5'-CGCCATGGTTAAGTCCTTATTTATTGC-3', RP-SodC-5'-CGCTCGAGTTCGATCAC GCCGCAGGCAAAA-3', respectively.

Amplification was performed in 50 µl containing 5 µl of 10 × PCR buffer (Qiagen), 1 µl of 10 mM dNTPs (New England Biolabs, USA), 0.1 µl of DNA, 1 µl of each primer (20 pMol/µl), 0.5 µl of Taq DNA polymerase (2.5 units, Qiagen). Amplification conditions: 94 °C 5 min; then 30 cycles of 94 °C, 1 min, 50 °C, 1 min, 72 °C, 2 min, and 1 cycle of 72 °C, 7 min.

The obtained products were digested with respective enzymes *NdeI* – *XhoI* (L7/L12), *NcoI* – *XhoI* (SodC) and cloned into the plasmid vector pET28b(+) (Novogen, USA).

As a result, recombinant plasmids pET/Br-L7/L12 and pET/Br-SodC were obtained containing the sequence coding the SodC protein fragment (1-173 aa) and the C-terminal peptide LEHHHHHH; and the sequence encoding the L7/L12 protein fragment (21-144 aa), the N-terminal peptide MSSHHHHHHSS and the C-terminal peptide LEHHHHHH. Plasmids were sequenced to verify the integrity of the inserts.

Plasmids were transformed into *E. coli* cells, strain T7 express (New England Biolabs, USA). As the result *E. coli* clones, producers of the recombinant proteins SodC and L7/L12, were obtained.

Gene expression. To expression the genes of the target proteins L7/L12 and SodC, bacterial cells were grown in LB-kan (Luria-Bertani broth, containing 50 µg/mL of kanamycin) at 37 °C on a shaker (250 rpm) to optical density OD600 = 0.6-1.0. Gene expression was induced by addition IPTG to final concentration 1 mM to the bacterial suspension with subsequent incubation for 4 h at 37 °C. The cells were harvested by centrifugation at 5000 × g for 15 min and stored at - 70 °C until further use. Aliquots

selected before and after induction were examined by PAGE electrophoresis. The solubility of the recombinant protein was determined using a BugBuster master mix reagent (Novagen, USA) according to the manufacturer's instructions.

Protein L7/L12 purification. Cell pellet was resuspended in NB buffer (50 mM NaH₂PO₄, 300 mM NaCl, 6 M guanidine hydrochloride, 10 mM imidazole, pH 7.4) at the rate of 5 ml of buffer per 1 g of crude cell pellet. Suspension was incubated in ice for 30 min and sonicated. Cell lysate was clarified by centrifugation at 3000 g for 15 min. The supernatant (soluble protein fraction) was filtered through a 0.22 µm membrane and uploaded to HisPur™ Cobalt Superflow (Thermo Scientific, USA) agarose pre-equilibrated with NB buffer. The agarose resin was washed using NW buffer (50 mM NaH₂PO₄, 300 mM NaCl, 20 mM imidazole, pH 7.4), the protein was eluted with NE buffer (50 mM NaH₂PO₄, 300 mM NaCl, 300 mM imidazole, pH 7.4). The pure protein fraction was dialyzed against 10 times the volume of the buffer (20 mM PBS, 300 mM NaCl, pH 7.4) overnight at 4°C.

Protein SodC purification. Protein purification was performed as [17] with modifications. Cells were resuspended in buffer 1 (100 mM Tris HCl pH 8.0, 150 mM NaCl, 1 % Triton X-100, 1 % Sodium deoxycholate) at the rate of 15 ml per 1 g of crude cell pellet. Lysozyme was added to the resulting suspension to a final concentration of 1 mg/ml. Cell lysis was performed by freezing twice at -70 °C and thawing at 37 °C of the suspension. The cell lysate was incubated at 4 °C overnight, then centrifuged at 4 °C for 15,000 × g for 15 min. The pellet was washed successively with buffer 2 (100 mM Tris HCl, pH 8.0, 150 mM NaCl, 1 % Triton X-100), buffer 3 (100 mM Tris HCl, pH 8.0, 1 M NaCl), buffer 4 (50 mM Tris HCl, pH 7.5). The pellet was resuspended in buffer 5 (20 mM PBS, pH 7.4, 300 mM NaCl, 8 M urea, 5 mM imidazole, pH 7.4) and incubated at 4°C overnight to completely solubilize. The dissolved inclusion fraction was filtered through a 0.22 µm membrane. Protein purification was carried out using HisPur™ Cobalt Superflow agarose. After sorption of protein, resin was washed with DW buffer (50 mM NaH₂PO₄, 300 mM NaCl, 8 M urea, 5 mM imidazole, pH 7.4). Protein was eluted with DE buffer (50 mM NaH₂PO₄, 300 mM NaCl, 8 M urea, 100 mM imidazole, pH 7.4). Refolding of the protein was performed by dialysis sequentially against buffer 7 (20 mM PBS, 300 mM NaCl, 4 M urea, pH 7.4), buffer 8 (20 mM PBS, 300 mM NaCl, 2 M urea, pH 7.4), buffer 9 (20 mM PBS, 300 mM NaCl, 1 mM DTT (dithiothreitol), pH 7.4) and buffer 10 (20 mM PBS, 300 mM NaCl, pH 7.4). Protein concentration was determined by the method of Lowry et al. [18], using BSA as a standard.

Polyacrylamide gel electrophoresis and Western blot. Electrophoretic analysis of polypeptides was performed in 12 % SDS-PAGE under denaturing reducing conditions according to Laemmli [19]. For visualization of proteins, Coomassie G-250 staining was used. For Western blot analysis, proteins were transferred onto a nitrocellulose membrane and detected as described in [20] using anti-His (Cterm)/AP antibodies (Invitrogen, USA) and sera from sick cattle.

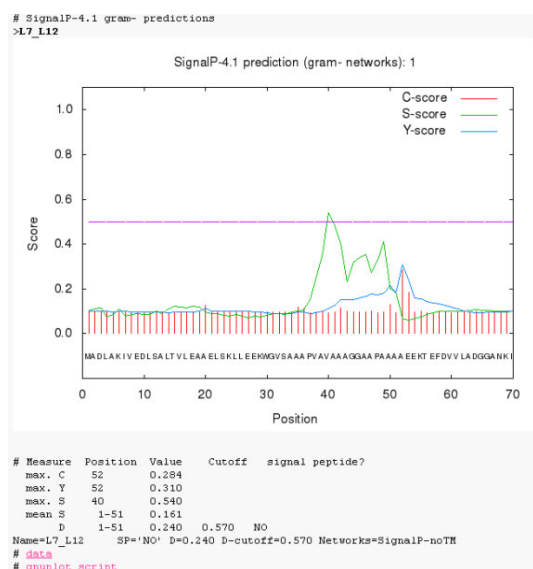
Obtaining specific serum to recombinant proteins. Animal experiments were carried out in accordance with applicable national and international legislation. The protocol was approved by the Bioethics Commission of the RIBSP RK ME&S of the Republic of Kazakhstan (No. 6 dated September 25, 2017).

To obtain specific sera, outbred white mice (females, 6-8 weeks old, weight 18-20 g) were immunized with a target protein. Proteins were prepared as follows: purified protein was mixed with Montanide Gel 01 (SEPPIC, USA) in a ratio of 9:1 (v/v). The final protein concentration was 150 µg/ml. Before administration of the drug, animals were bled to obtain normal serum. Immunization was carried out subcutaneously four times in a dose of 30 µg of protein. 14 days after the last injection, the animals were bled. The serum was tested in ELISA (enzyme-linked immunosorbent assay).

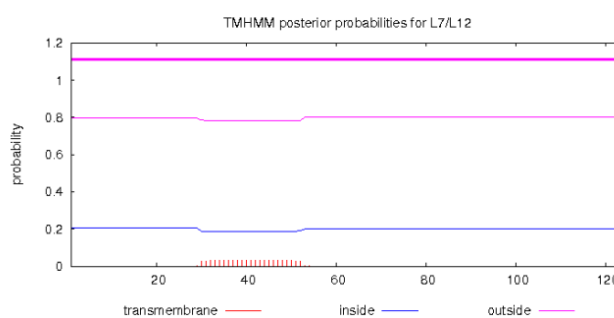
ELISA. 96-wells plates were coated with the 2 mg/ml of affinity purified recombinant proteins in 0.05 M carbonate-bicarbonate buffer, pH 9.6 (100 µL/well), and incubated overnight at 4 °C. Plates were washed four times with TBST buffer (150 mM NaCl, 20 mM tris-HCl, pH 7.5, 0.1 % tween-20) and blocked with TBST containing 5 % fat free dry milk for 1 h at 37 °C. Double dilutions of test sera in the blocking buffer, were added to wells (100 µL/well). Plates were incubated for 1 h at 37 °C and washed three times with TBST. Anti-mouse immunoglobulin IgG conjugated to alkaline phosphatase (1:5000) was added (100 µL/well) and the plates were incubated for 1 h at 37 °C. After washing, the substrate for alkaline phosphatase (pNPP, Sigma, USA) was added into each well (100 µL). The plates were incubated

for 30 min. Optical density was read at 405/630 nm on ELISA plates reader ImmunoChem-2100. Cut-off values were determined using the mean optical density values from negative control sera plus three standard deviations.

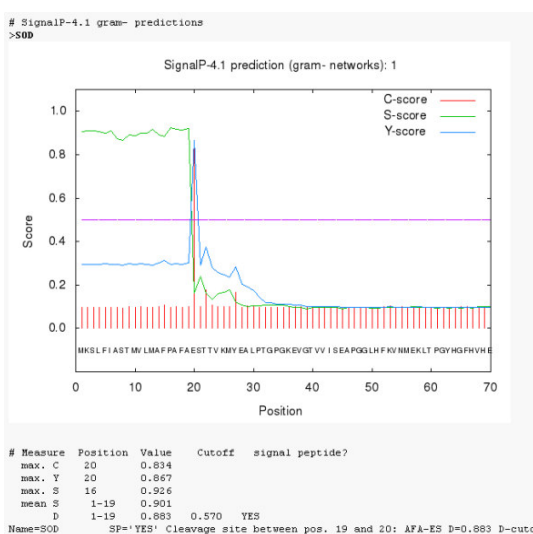
Results and discussion. *Construction of recombinant plasmids.* A comparative analysis of the amino acid sequences of the L7/L12 and SodC proteins showed their high identity (95-100 %) for *Brucella spp.* Using the SignalP [21] and TMHMM2.0 [22] software, the signal peptide was established for the SodC protein, while transmembrane domains were absent in both proteins (figure 1).



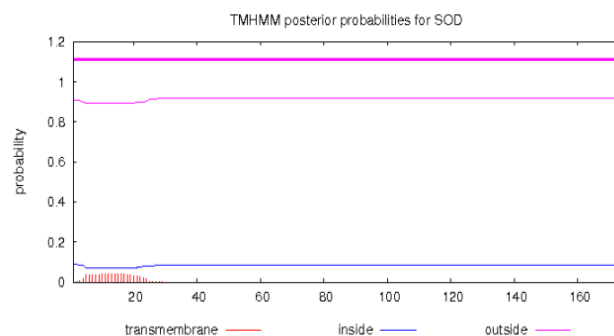
```
# L7/L12 Length: 124
# L7/L12 Number of predicted TMs: 0
# L7/L12 Exp number of AAs in TMs: 0.73504
# L7/L12 Exp number, first 60 AAs: 0.73504
# L7/L12 Total prob of N-in: 0.20413
L7/L12 TMHMM2.0 outside 1 124
```



A



```
# SOD Length: 173
# SOD Number of predicted TMs: 0
# SOD Exp number of AAs in TMs: 0.74997
# SOD Exp number, first 60 AAs: 0.74997
# SOD Total prob of N-in: 0.09191
SOD TMHMM2.0 outside 1 173
```



B

Figure 1 – The results of the analysis of the amino acid sequences of the target proteins for the presence of signal peptides and transmembrane domains: A – L7/L12, B – SodC

Amplified DNA fragments encoding the L7/L12 and SodC genes were cloned into the pET28b(+) vector. As a result, plasmids expressing recombinant proteins flanked at the N- and/or C-terminus by 6HIS oligopeptides were obtained (figure 2). The predicted molecular weights of SodC and L7/L12 proteins were 19.87 and 15.77 kDa, respectively. The obtained plasmids were transformed into *E. coli* T7 cells.

Analysis	Entire Protein
Length	152 aa
Molecular Weight	15774.20
1 microgram =	63.395 pMoles
Molar Extinction coefficient	5690
1 A[280] corr. to	2.77 mg/ml
A[280] of 1 mg/ml	0.36 AU
Isoelectric Point	6.10
Charge at pH 7	-5.07

1	MGSSHHHHHH	SSGLVPRGSH	MADLAKIVED	LSALTVLEAA	ELSKLLEEKW
51	GVSAAAPVAV	AAAGGAAPAA	AAEKEFEFV	VLADGGANKI	NVIKEVRALT
101	GLGLKEAKDL	VEGAPKAVKE	GASKDEAEKI	KAQLEAAGAK	VELKLEHHHH
151	HH				

Analysis	Entire Protein
Length	188 aa
Molecular Weight	19875.26
1 microgram =	50.314 pMoles
Molar Extinction coefficient	5360
1 A[280] corr. to	3.71 mg/ml
A[280] of 1 mg/ml	0.27 AU
Isoelectric Point	6.37
Charge at pH 7	-3.75

1	MKSLFIASMTM	VLMAFFPAF&E	STTVKMYEAL	PTGPGKEVGT	VVISEAPGGL
51	HFKNMHEKLT	PGYHGPHVHE	NPSC&PGEKD	GKIVPALAAG	GHYDPGNTHH
101	HLGPEGDGHH	GDLPELSANA	DGKVS&TVVA	PHLKKLAEIK	QRSLMVHVGG
151	DNYS&RPEPL	GGGARF&C	VIEDKLA&AL	EH&HHHHH*	

A

B

Figure 2 – Prediction and analysis of amino acid sequences of recombinant proteins L7/L12 (A) and SodC (B)

The expression of genes encoding recombinant proteins. Induction of target genes expression resulted in the production of L7/L12 (figure 3A) and SodC (Figure 3B) proteins. The molecular weight of the recombinant proteins corresponded to the calculated values (figures 2, 3). As a result of gene SodC expression, two protein products were formed (figure 3B, Tot), which is associated with the presence of a signal peptide in the sequence of the recombinant protein. Modified protein (without signal peptide) was in the soluble protein fraction (figure 3B, So), while unmodified protein formed inclusions bodies (figure 3B, IN).

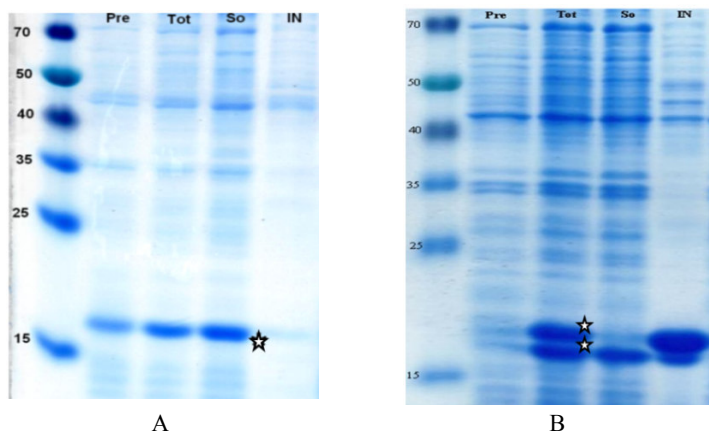


Figure 3 –

Electrophoretic analysis of proteins of the cell lysate of *E. coli* strain T7, transformed with plasmids pL7/L12 (A) and pSodC (B): Pre – cell lysate before induction, Tot – after induction IPTG, So – soluble proteins, IN – inclusions. Recombinant proteins are marked with asterisks

The expression of the target *Brucella* L7/L12 and SodC proteins was confirmed by the Western blot using anti-His-antibodies (figure 4). Recombinant proteins also interacted with sera from brucellosis sick cattle, which confirms their specificity (figure 5).

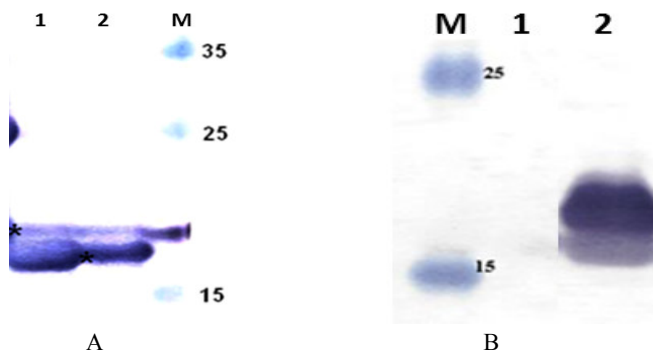
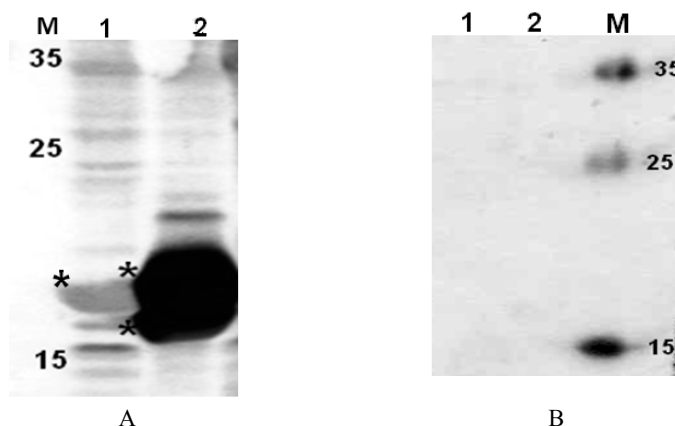


Figure 4 –

Immunoblotting of proteins of the cell lysate of *E. coli* strain T7, transformed with recombinant pET plasmids, using serum to polyhistidine. A: 1 – cell lysate prior to induction of L7/L12; 2 – after induction of IPTG; B: 1 – cell lysate prior to SodC induction; 2 – after induction of IPTG

Figure 5 –
Immunoblotting of proteins of the cell lysate of *E. coli* strain T7, transformed with recombinant pET plasmids, using serum from cattle with brucellosis:
A – serum from cattle with brucellosis,
B – normal cattle serum.
1 – cell lysate L7/L12;
2 – cell lysate SodC. Recombinant proteins are marked with asterisks



B. abortus Cu-Zn супероксиддисмутаза (SodC) была идентифицирована Beck et al. (1990) [23]. Using western blot, Betsy et al. (1990) proved that superoxide dismutase is found in most *Brucella* strains and species except *B. neotomae* and *B. suis* biovar 2 [24]. Both SodC and L7/L12 are immunodominant proteins and induce antibody production. Rajagunalan et al. (2014) in their studies, found antibodies to the recombinant protein L7/L12 of *B. melitensis* 16M in the sera from patients with acute brucellosis [25]. Proteins SodC and L7 / L12 were used in the development of DNA and vector vaccines. Recombinant vaccines elicited an immune response in animals [13, 14, 26].

Protein purification L7/L12. At the first stage, the purification of L7/L12 protein was carried out under native conditions (figure 6).

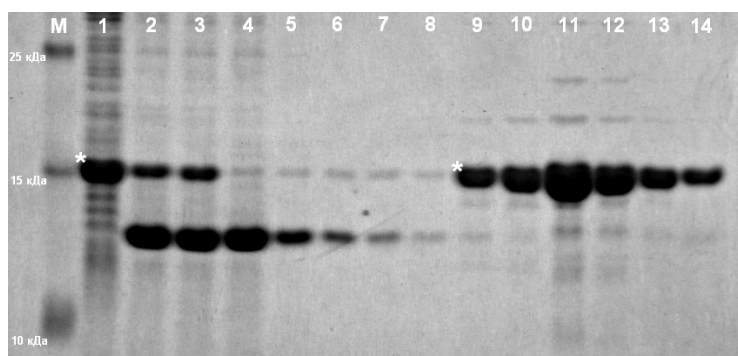


Figure 6 – Electrophoretic analysis of the recombinant protein L7/L12 during the cleaning process:
M – molecular weight marker; 1 – total cell lysate; 2 – cell lysate after treatment with lysozyme;
3 – cell lysate after filtration through 0.22 μm ; 4 – flow-through; 5-8 – wash fraction;
9-14 – fractions of purified protein. Recombinant proteins are marked with asterisks

As seen in the figure 6, lane 4, the cell lysate was not completely adsorbed onto resin. Washing the agarose with a buffer containing both 20 mM imidazole (figure 6, lane 7, 8) and 10 mM imidazole (figure 6, lane 5, 6) resulted in the loss of the target protein. The target protein eluted with a buffer containing both 300 mM imidazole (figure 6, lane 11-14) and 100 mM imidazole (figure 6, lane 9, 10) contained cell protein impurities. Purification of the protein under standard native conditions according to the recommendations of the resin manufacturer did not give satisfactory results. The protein purification protocol was optimized.

Hybrid conditions were selected for purification (see the Materials and Methods section), which made it possible to obtain a protein preparation with a high degree of purification without significant losses at intermediate stages (figure 7). The yield of recombinant protein L7/L12 was 13 mg per 1 liter of culture.

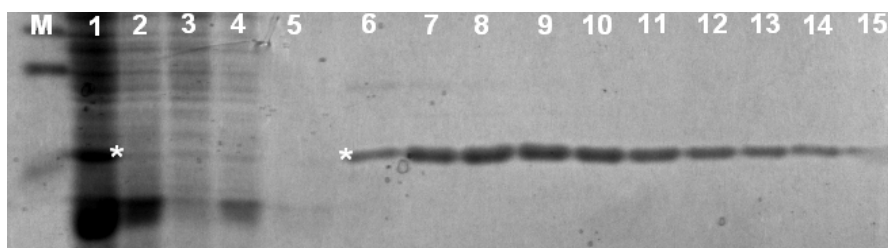


Figure 7 – Electrophoretic analysis of the recombinant protein L7/L12 during the purification process using optimal (hybrid) conditions. M – molecular weight marker; 1 – cell lysate after treatment with lysozyme; 2 – flow-through; 3-5 – washing fractions; 6-15 – fractions of purified protein. Recombinant proteins are marked with asterisks

SodC Protein Purification. Protein purification was performed under denaturing conditions (figure 8). When purifying the recombinant SodC protein, there were also problems with its adsorption to agarose. Only a small amount of protein was bound to the resin (figure 8, lane 2). Washing the resin led to the loss of the target protein (figure 8, lane 3-6), and the protein eluate contained a significant amount of impurities of cellular proteins (figure 8, lanes 7-10). An optimized purification protocol (see the Materials and Methods section) made it possible to obtain a SodC protein preparation with a high degree of purity (figure 9).

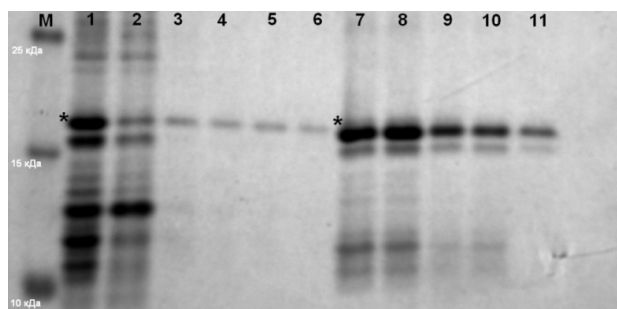


Figure 8 – Electrophoretic analysis of recombinant SodC protein during purification under denaturing conditions. M – molecular weight marker; 1 – cell lysate after treatment with lysozyme; 2 – flow-through; 3-6 – wash fraction; 7-12 – fractions of purified protein. Recombinant proteins are marked with asterisks

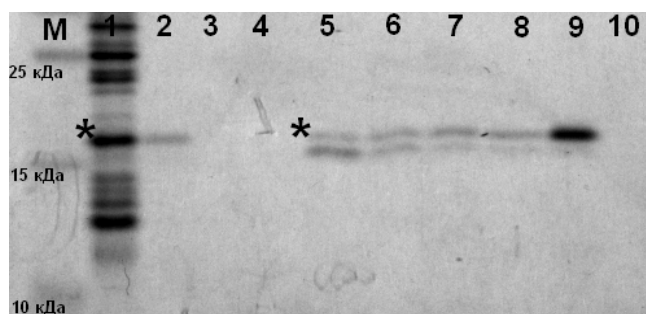


Figure 9 – Electrophoretic analysis of recombinant SodC protein during purification under optimal denaturing conditions: M – molecular weight marker; 1 – cell lysate after treatment with lysozyme; 2 – flow-through; 3-4 – washing fractions; 5-10 – fractions of purified protein. Recombinant proteins are marked with asterisks

As seen in the figure 9, a slight loss of the target protein was observed upon binding to agarose (lane 2). Loss of the target protein during resin washing was not detected (lanes 3-4). There were no impurities of cell protein in the eluate (lanes 5-9), which confirms the high purity of the obtained recombinant protein preparation. The yield of purified SodC protein was 10 mg per 1 liter of culture.

In addition, mouse specific sera for recombinant SodC and L7 / L12 proteins were obtained. For this, mice were immunized with triply recombinant proteins mixed with Montanide 01 gel (seppic, USA). Recombinant proteins induced the production of specific antibodies in animals. The antibody titer in ELISA for both SodC and L7 / L12 proteins was 1: 20480.

Purified recombinant proteins and their specific sera are suitable for use in the development of specific diagnostic and prophylactic agents against *Brucella* spp.

Conclusion. As a result of studies, purified preparations of recombinant proteins SodC and L7/L12 were obtained. Optimal protocols have been developed for the expression and purification of recombinant proteins in E.coli. The specificity of the obtained purified recombinant proteins was established using the western blot with sera from animals with brucellosis. Highly active specific sera for recombinant proteins were obtained, an antibody titer was 1:20480 for both the SodC and the L7/L12 proteins. Proteins and

serums for them will be further used in the development of specific diagnostic and prophylactic agents against *Brucella* spp.

Acknowledgments. This work was supported by the Ministry of Education and Science of the Republic of Kazakhstan (grant No. AP05133746) for 2018-2020.

Э. Т. Тайлакова, С. О. Садикалиева, Г. О. Шыныбекова,
А. К. Абубакирова, К. Т. Султанкулова, О. В. Червякова

Биологиялық қауіпсіздік проблемаларын ғылыми зерттеу институты,
Гвардейский, Қордай ауданы, Жамбыл облысы, Қазақстан

***E. COLI*-ДЕ *BRUCELLA* *SPP.* L7/L12 ЖӘНЕ SodC РЕКОМБИНАНТТЫ БЕЛОКТАРДЫ КОНСТРУИРЛЕУ, ЭКСПРЕССИЯЛАУ ЖӘНЕ ТАЗАЛАУ**

Аннотация. Бруцеллез инфекцияның табиғи резервуарлары бар болғандықтан, денсаулық сақтаудың маңызды проблемасы болып қала береді. Қазақстанның жеке аудандарында жыл сайын адамдардың және ауыл шаруашылығындағы малдардан бруцеллездің анықталуы індет бойынша тұрақсыз, құбылмалы жағдайды және шаруашылықтардағы эпизоотиялық және эпидемиялық процестердің әртүрлі деңгейдегі белсенді көріністерімен бруцеллез ошақтарының қалыптастыруының нақты мүмкіндігінің бар екенін білдіреді. Осыған байланысты, бұл індет ветеринария және медицина ғылымы үшін үлкен мәселе болып отыр. Қазіргі таңда бруцеллезді мал шаруашылығында алдын алу үшін S19 және RB51 штамдары негізінде алынған тірі аттенуирленген вакциналар қолданылады. Олардың басты кемшілігі – адамдарға тигізетін жұқпалы әсері. Сонда да адамдардың бұл індетпен ауруын төмендету үшін ауыл шаруашылығындағы малдарды иммундау бағдарламалары жүзеге асырылуы қажет. Омиксті технологияларды қолдана отырып, қауіпсіз және тиімді болатын жаңа дәуір вакциналарын жасақтау – вакцинологияның перспективті бағыты. Бүгінгі күні алдын алу және балау, препараттарды әзірлеу кезінде, сонымен қатар әртүрлі зоонозды ауруларға қарсы вакцина жасау барысында қолданылатын, антигендік және иммуногендік қасиеттері бар рекомбинантты белоктар кеңінен қолданылады. Гуморалды және жасуша деңгейінде иммунды жауап қайтаратын бруцелланың бірқатар иммуногенді белоктары айқындалған болатын. Бұл мақаланы жазудың мақсаты *Brucella* spp. L7/L12 және SodC рекомбинантты белоктарды экспрессиялау және тазалау барысындағы жағдайларды оңтайландырумен байланысты. Жасалған жұмыстардың нәтижесінде экспрессиялайтын плазмидалық ДНҚ *pET/Br-L7/L12* және *pET/Br-SodC* алынды. Тұтас гендердің *E.coli* -де экспрессиялау параметрлері анықталды және рекомбинантты белоктарды тазалау әдісі оңтайландырылды. Нәтижесінде L7/L12 белокты тазалау үшін келесідей гибридті жағдай таңдалды: жасушаларды лизистеу барысында құрамында 6М гуанидин гидрохлориді бар буферді қолдану, жуып-шаю барысында құрамында 20 мМ имидазол бар буферді қолдану, HisPur агарозадан тұтас белокты элюирлеу барысында, құрамында 300 мМ имидазол бар буферді қолдану. SodC белокты денатурациялы жағдайда лизис буферіне 1 % тритон X-100 және 1 % натрий дезоксихолаты қосылып тазаланды. Алынған тұнбалар құрамында 8 М мочевина және 5 мМ имидазол бар буферді қолдана отырып, ерітіліп алынды. HisPur агарозадан тұтас белокты құрамында 8 М мочевина және 100 мМ имидазол бар буферді қолдана отырып, элюирлеп алынды. Тазалау барысының өзгертілген жағдайларын қолдана отырып, тазартылған рекомбинантты белоктар келесідей мөлшерде алынды: L7/L12 шығымдылығы 13 мг/л, SodC шығымдылығы 10 мг/л. Белоктардың телімділігі вестр блотты қолдана отырып расталды. Рекомбинантты белоктармен тышқандарды егу нәтижесінде ИФА-де титрлері 1:20480 және 1:20480 сәйкес телімді антиденелердің пайда болуы анықталды. Рекомбинантты белоктар мен оларға алынған сарысулар бруцеллезге қарсы қой шешегі вирусы негізде векторлық вакцинаны жасауда қолданылатын болады.

Түйін сөздер: *Brucella* spp., рибосомалық белок L7/L12, супероксиддисмутаза, экспрессия, тазалау.

Э. Т. Тайлакова, С. О. Садикалиева, Г. О. Шыныбекова,
А. К. Абубакирова, К. Т. Султанкулова, О. В. Червякова.

Научно-исследовательский институт проблем биологической безопасности
пгт. Гвардейский, Кордайский район, Жамбылская область, Казахстан

КОНСТРУИРОВАНИЕ, ЭКСПРЕССИЯ И ОЧИСТКА РЕКОМБИНАНТНЫХ БЕЛКОВ L7/L12 И SODC *BRUCELLA SPP.* В *E. COLI*

Аннотация. Бруцеллез продолжает оставаться важной проблемой здравоохранения, пока существуют естественные резервуары инфекции. Ежегодное выявление реагирующих на бруцеллез сельскохозяйственных животных и людей в отдельных районах Казахстана свидетельствует о крайне неустойчивой ситуации по этой инфекции и о реальной возможности формирования очагов бруцеллеза с разной степенью активности проявления эпизоотических и эпидемических процессов в хозяйствах. В связи с этим данная инфекция представляет серьезнейшую проблему для ветеринарной и медицинской науки. В настоящее время для профилактики бруцеллеза у животных используют живые аттенуированные вакцины на основе штаммов S19 и RB51, главным недостатком которых является вирулентность для человека. Тем не менее, программы иммунизации животных должны проводиться, чтобы снизить заболеваемость людей. Разработка безопасных и эффективных вакцин нового поколения с использованием омиксных технологий является перспективным направлением вакцинологии. Сегодня при разработке профилактических и диагностических препаратов, а также в качестве кандидатов на вакцину против различных зоонозных заболеваний широко используются рекомбинантные белки, обладающие антигенными и иммуногенными свойствами. Установлен ряд иммуногенных белков бруцелл, индуцирующих как гуморальный, так и клеточный иммунный ответ. Целью данных исследований являлась оптимизация условий экспрессии и очистки рекомбинантных белков L7/L12 и SodC *Brucella spp.* В результате проведенных исследований получены экспрессирующие плазмидные ДНК *pET/Br-L7/L12* и *pET/Br-SodC*. Установлены параметры экспрессии целевых генов в *E.coli* и оптимизирован метод очистки рекомбинантных белков. В результате проведенных работ для очистки белка L7/L12 подобраны гибридные условия с использованием буфера содержащего 6М гуанидина гидрохлорида на этапе лизиса клеток, 20 мМ имидазола на этапе отмывки и 300 мМ имидазола на этапе элюирования целевого белка с агарозы HisPur. Белок SodC очищали в денатурирующих условия с добавлением в лизирующий буфер 1 % тритон X-100 и 1 % дезоксихолата натрия. Включения сольubilizировали буфером, содержащим 8 М мочевины и 5 мМ имидазола. Элюировали целевой белок с агарозы HisPur буфером содержащим 8 М мочевины и 100 мМ имидазола. Использование модифицированных протоколов очистки позволило получить очищенные рекомбинантные белки с выходом 13 мг/л для белка L7/L12 и 10 мг/л для белка SodC, соответственно. Специфичность белков была подтверждена в вестерн блоте. Иммунизация мышей рекомбинантными белками приводила к выработке специфических антител, титр которых в ИФА составил 1:20480 и 1:20480 соответственно. Рекомбинантные белки и специфические сыворотки к ним будут использованы при разработке векторной противобруцеллезной вакцины на основе вируса оспы овец.

Ключевые слова: *Brucella spp.*, рибосомальный белок L7/L12, супероксиддисмутаза, экспрессия, очистка.

Information about authors:

Tailakova E.T., master of biology, researcher of the laboratory of “Molecular biology and genetic engineering”, Research Institute for Biological Safety Problems, Korday district, Zhambyl region, Gvardeiskiy v., Kazakhstan; tailakova_86@mail.ru; <https://orcid.org/0000-0003-3987-7451>

Sadikaliyeva S.O., master of chemistry, researcher of the laboratory of “Molecular biology and genetic engineering”, Research Institute for Biological Safety Problems, Korday district, Zhambyl region, Gvardeiskiy v., Kazakhstan; sadikaliyeva86@mail.ru; <https://orcid.org/0000-0003-4953-3628>

Shynybekova G.O., master of biology, senior assistant of the laboratory of “Molecular biology and genetic engineering”, Research Institute for Biological Safety Problems, Korday district, Zhambyl region, Gvardeiskiy v., Kazakhstan; gaukhar_1988@bk.ru; <https://orcid.org/0000-0002-6976-1540>

Abubakirova A.K., master of veterinary sciences, senior assistant of the laboratory of “Molecular biology and genetic engineering”, Research Institute for Biological Safety Problems, Korday district, Zhambyl region, Gvardeiskiy v., Kazakhstan; aigerim_abubakirova94@mail.ru; <https://orcid.org/0000-0003-3601-8977>

Sultankulova K.T., candidate of biological sciences, professor, head of the laboratory of “Molecular biology and genetic engineering”, Research Institute for Biological Safety Problems, Korday district, Zhambyl region, Gvardeiskiy v., Kazakhstan; sultankul70@mail.ru; <https://orcid.org/0000-0002-1332-1247>

Chervyakova O.V., candidate of biological sciences, leading researcher of the laboratory of “Molecular biology and genetic engineering”, Research Institute for Biological Safety Problems, Korday district, Zhambyl region, Gvardeiskiy v., Kazakhstan; ovch@mail.ru; <https://orcid.org/0000-0002-7954-6246>

REFERENCES

[1] Ivanov N.P. (2013) Diseases of ruminants, pigs and horses, diseases of birds, carnivores and fur-bearing animals, bees, fish, little-known diseases and slow infections [Bolezni zhvachnyh zhivotnyh, svinej i loshadej, bolezni ptic, plotojadnyh i pushnyh zverej, pchel, ryb, maloizvestnye bolezni i medlennye infekcii]. Infectious animal diseases: tutorial [Infektsionnye bolezni zhivotnykh: uchebnoe posobie], Almaty: Nur-print. ISBN: 978-601-241-370-0 (in Russ.).

[2] Ivanov N.P. (2007) Animal brucellosis and control measures: tutorial [Brutsellez zhivotnykh i mery borby s nim: uchebnoe posobie], Almaty: Atamura (in Russ.).

[3] Janyshv A.A. (2007) Cattle and small brucellosis in the Southern Federal District: epizootological monitoring and improvement of anti-epizootic measures [Brucelljoz krupnogo i melkogo rogatogo skota v Juzhnom Federal'nom okruge: epizootologicheskij monitoring i sovershenstvovanie protivoevizooticheskikh meroprijatij], Avtoref. dis. k.v.n., M., 21 p. (in Russ.).

[4] Enright F.M., Nielsen K. and Duncan J.R. (1990) The pathogenesis and pathobiology of *Brucella* infection in domestic animals. Animal Brucellosis, CRC Press, Boca Raton, FL, P. 301–320. ISBN:0849358787 (in Eng.).

[5] Sattarov A.I. (2005) Brucellosis in the Republic of Kazakhstan, Materials of the international scientific-practical conference dedicated to the 100th anniversary of KazNIVI [Brucellez v Respublike Kazahstan, Materialy mezhdunarodnoj nauchno-prakticheskoy konferencii, posvjashhennoj 100 – letiju KazNIVI], Almaty, P. 237-241 (in Russ.).

[6] Ivanov N.P. (2014) Special veterinary measures for brucellosis of farm animals in the Republic of Kazakhstan [Special'nye veterinarnye meroprijatija pri brucelleze sel'skohozjajstvennyh zhivotnyh v Respublike Kazahstan], *Veterinary medicine* [Veterinarija], 1(35): 18-21 (in Russ.).

[7] Pasquevich K.A., Ibanez A.E., Coria L.M., García Samartino C., Estein S.M., Zwerdling A., Barrionuevo P. (2011) An oral vaccine based on U-Omp19 induces protection against B. abortus mucosal challenge by inducing an adaptive IL-17 immune response in mice. *PLoS One*, 6 (1). <https://doi.org/10.1371/journal.pone.0016203> (in Eng.).

[8] Saez D., Guzman I., Andrews E., Cabrera A., Onate A. (2008) Evaluation of *Brucella abortus* DNA and RNA vaccines expressing Cu-Zn superoxide dismutase (SOD) gene in cattle. *Veterinary microbiology*, 129(3): 396-403. <https://doi:10.1016/j.vetmic.2007.11.015> (in Eng.).

[9] Commander N.J., Spencer S.A., Wren B.W., MacMillan A.P. (2007) The identification of two protective DNA vaccines from a panel of five plasmid constructs encoding *Brucella melitensis* 16M genes. *Vaccine*, 25(1): 43-54. <https://doi:10.1016/j.vaccine.2006.07.046> (in Eng.).

[10] Bruno C.A., Estein L., Barrera S.M., Bowden S., Fossati R. (2007) Improved immunogenicity of a vaccination regimen combining a DNA vaccine encoding *Brucella melitensis* outer membrane protein 31 (Omp31) and recombinant Omp31 boosting. *Clinical and vaccine immunology*, 14(7): 869-874. <https://doi:10.1128/CVI.00472-06> (in Eng.).

[11] Pasquevich K.A., Estein S.M., Garcia Samartino C., Zwerdling A., Coria L.M., Barrionuevo P., Fossati C.A., Giambartolomei G.H., Cassataro J. (2008) Immunization with recombinant *Brucella* spp. outer membrane proteins Omp16 or Omp19 in adjuvant induces specific CD41 and CD81 T cells as well as systemic and oral protection against *Brucella abortus* infection. *Infection Immunity*, 77: 436-445. <https://doi:10.1128/IAI.01151-08> (in Eng.).

[12] Yu D.H., Hu X.D., Cai H.A. (2007) Combined DNA vaccine encoding BCSP31, SOD, and L7/L12 confers high protection against *Brucella abortus* 2308 by inducing specific CTL responses. *DNA. Cell Biology*, 26: 435-43. <https://doi:10.1089/dna.2006.0552> (in Eng.).

[13] Luo D., Ni B., Li P. (2006) Protective immunity elicited by a divalent DNA vaccine encoding both the L7/L12 and Omp16 genes of *Brucella abortus* in BALB/c mice. *Infection Immunity*, 74: 2734-41. <https://doi:10.1128/IAI.74.5.2734-2741.2006> (in Eng.).

- [14] Kurar E., Splitter G.A. (1997) Nucleic acid vaccination of *Brucella abortus* ribosomal L7/L12 gene elicits immune response. *Vaccine*, 15: 1851-1857. [https://doi:10.1016/s0264-410x\(97\)00140-0](https://doi:10.1016/s0264-410x(97)00140-0) (in Eng.).
- [15] Zhao Z., Li M., Luo D. (2009) Protection of mice from *Brucella* infection by immunization with attenuated *Salmonella enterica* serovar typhimurium expressing A L7/L12 and BLS fusion antigen of *Brucella*. *Vaccine*, 27: 5214-5219. <https://doi:10.1016/j.vaccine.2009.06.075> (in Eng.).
- [16] Munoz-Montesino C., Andrews E., Rivers R., Gonzalez-Smith A., Moraga-Cid G., Folch H., Cespedes S., Onate A.A. (2004) Intraspleen delivery of a DNA vaccine coding for superoxide dismutase (SOD) of *Brucella abortus* induces SOD-specific CD4(1) and CD8(1) T cells. *Infection Immunity*, 72: 2081-2087. <https://doi:10.1128/iai.72.4.2081-2087.2004> (in Eng.).
- [17] Singh K., Gittis A.G., Gitti R.K., Ostazeski S.A., Su H-P., Garboczi D.N. (2016) The vaccinia virus H3 envelope protein, a major target of neutralizing antibodies, exhibits a glycosyltransferase fold and binds UDP-glucose. *Journal Virology*, 90: 5020-5030. <https://doi:10.1128/JVI.02933-15> (in Eng.).
- [18] Lowry O.H., Rosebraugh N.J. (1951) Protein measurement with the Folin phenol reagent. *Journal Biology Chemistry*, 193: 265-275. PMID: 14907713 (in Eng.).
- [19] Laemmli U.K. (1970) Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227: 680-685. <https://doi.org/10.1038/227680a0> (in Eng.).
- [20] Tailakova E.T., Chervyakova O.V. (2016) Bacterial gene expression of sheep pox virus encoding antigenic proteins of SPPV095 and SPPV141 for the development of new generation specific prophylaxis. *Biotechnology. Theory and practice*, 2: 81-87 (in Eng.).
- [21] Moller S., Croning M.D.R., Apweiler R. (2001) Evaluation of methods for the prediction of membrane spanning regions. *Bioinformatics*, 17(7): 646-653. <https://doi:10.1093/bioinformatics/17.7.646> (in Eng.).
- [22] Armenteros J.J.A., Tsirigos K.D., Sonderby C.K., Petersen T.N., Brunak O.W.S., Heijne G., Nielsen H. (2019) SignalP 5.0 improves signal peptide predictions using deep neural networks. *Nature Biotechnology*, 37: 420-423. <https://doi:10.1038/s41587-019-0036-z> (in Eng.).
- [23] Beck BL, Tabatabai LB, Mayfield JE (1990) A protein isolated from *Brucella abortus* is a Cu-Zn superoxide dismutase. *Biochemistry*, 29: 372-376. <https://doi:10.1021/bi00454a010> (in Eng.).
- [24] Bricker B.J., Tabatabai L.B., Judge B.A., Deyoe B.L., Mayfield J.E. (1990) Cloning, Expression, and occurrence of The *Brucella* Cu-Zn Superoxide dismutase. *Infection and Immunity*, 58(9): 2935-2939. PMID: 2201639 (in Eng.).
- [25] Rajagunalan S., Doimari S., Singh D.K. (2014) Cloning, Expression and Purification of L7/L12 Ribosomal Gene of *Brucella melitensis* 16M. *Journal of pure and applied microbiology*, 8(1): 1-6.
- [26] Tabynov K.K., Sansyzbay A.R., Kydyrbayev Zh.K., Yespembetov B.A., Ryskeldinova Sh.Zh., Zinina N.N., Assanzhanova N.N. et al. (2014) Influenza viral vectors expressing the *Brucella* OMP16 or L7/L12 proteins as vaccines against *B. abortus* infection. *Virology Journal*, 11: 69. <https://doi:10.1186/1743-422X-11-69> (in Eng.).

Publication Ethics and Publication Malpractice in the journals of the National Academy of Sciences of the Republic of Kazakhstan

For information on Ethics in publishing and Ethical guidelines for journal publication see <http://www.elsevier.com/publishingethics> and <http://www.elsevier.com/journal-authors/ethics>.

Submission of an article to the National Academy of Sciences of the Republic of Kazakhstan implies that the described work has not been published previously (except in the form of an abstract or as part of a published lecture or academic thesis or as an electronic preprint, see <http://www.elsevier.com/postingpolicy>), that it is not under consideration for publication elsewhere, that its publication is approved by all authors and tacitly or explicitly by the responsible authorities where the work was carried out, and that, if accepted, it will not be published elsewhere in the same form, in English or in any other language, including electronically without the written consent of the copyright-holder. In particular, translations into English of papers already published in another language are not accepted.

No other forms of scientific misconduct are allowed, such as plagiarism, falsification, fraudulent data, incorrect interpretation of other works, incorrect citations, etc. The National Academy of Sciences of the Republic of Kazakhstan follows the Code of Conduct of the Committee on Publication Ethics (COPE), and follows the COPE Flowcharts for Resolving Cases of Suspected Misconduct (http://publicationethics.org/files/u2/New_Code.pdf). To verify originality, your article may be checked by the Cross Check originality detection service <http://www.elsevier.com/editors/plagdetect>.

The authors are obliged to participate in peer review process and be ready to provide corrections, clarifications, retractions and apologies when needed. All authors of a paper should have significantly contributed to the research.

The reviewers should provide objective judgments and should point out relevant published works which are not yet cited. Reviewed articles should be treated confidentially. The reviewers will be chosen in such a way that there is no conflict of interests with respect to the research, the authors and/or the research funders.

The editors have complete responsibility and authority to reject or accept a paper, and they will only accept a paper when reasonably certain. They will preserve anonymity of reviewers and promote publication of corrections, clarifications, retractions and apologies when needed. The acceptance of a paper automatically implies the copyright transfer to the National Academy of Sciences of the Republic of Kazakhstan.

The Editorial Board of the National Academy of Sciences of the Republic of Kazakhstan will monitor and safeguard publishing ethics.

Правила оформления статьи для публикации в журнале смотреть на сайте:

www.nauka-nanrk.kz

ISSN 2518-1629 (Online), ISSN 2224-5308 (Print)

<http://biological-medical.kz/index.php/en/>

Редакторы: *М. С. Ахметова, Г. Б. Халидуллаева, Д. С. Аленов*
Верстка на компьютере *Д. А. Абдрахимовой*

Подписано в печать 02.04.2020.

Формат 60x881/8. Бумага офсетная. Печать – ризограф.
5,5 п.л. Тираж 300. Заказ 2.